

The Potential of Menstrual Blood-Derived Stem Cells in Differentiation to Epidermal Lineage: A Preliminary Report

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ABSTRACT

BACKGROUND

Menstrual blood-derived stem cells (MenSCs) are a novel source of stem cells that can be easily isolated non-invasively from female volunteered donor without ethical consideration. These mesenchymal-like stem cells have high rate of proliferation and possess multi lineage differentiation potency. This study was undertaken to isolate the MenSCs and assess their potential in differentiation into epidermal lineage.

METHODS

About 5-10 ml of menstrual blood (MB) was collected using sterile Diva cups inserted into vagina during menstruation from volunteered healthy fertile women aged between 22-30 years. MB was transferred into Falcon tubes containing phosphate buffered saline (PBS) without Ca²⁺ or Mg²⁺ supplemented with 2.5 µg/ml fungizone, 100 µg/mL streptomycin, 100 U/mL penicillin and 0.5 mM EDTA. Mononuclear cells were separated using Ficoll-Hypaque density gradient centrifugation and washed out in PBS. The cell pellet was suspended in DMEM-F12 medium supplemented with 10% FBS and cultured in tissue culture plates. The isolated cells were co-cultured with keratinocytes derived from the foreskin of healthy newborn male aged 2-10 months who was a candidate for circumcision for differentiation into epidermal lineage.

RESULTS

The isolated MenSCs were adhered to the plate and exhibited spindle-shaped morphology. Flow cytometric analysis revealed the expression of mesenchymal markers of CD10, CD29, CD73, and CD105 and lack of hematopoietic stem cells markers. An early success in derivation of epidermal lineage from MenSCs was visible.

CONCLUSION

The MenSCs are a real source to design differentiation to epidermal cells that can be used non-invasively in various dermatological lesions and diseases.

KEYWORDS

Menstrual blood-derived stem cells; Differentiation; Epidermal lineage

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INTRODUCTION

Stem cells as self-renewing cells proliferating without differentiation, and under defined conditions can differentiate into various cell types.¹ The stem cells can be categorized into two major groups of embryonic stem cells (ESCs) and adult stem cells (ASCs).² Embryonic stem cells are derived from inner cell mass of the blastocysts and show pluripotency characteristics by differentiation into all cells types belong to the three germinative layers (ectoderm, mesoderm, and endoderm).³ On the other hand, the ASCs are deposited in most adult tissues and are long-lived with restricted differentiation potency (Mehrabani et al. 2015),⁴ and lack tumorigenicity and ethical issues seen with ESCs.⁵

Friedensten et al (1968) were the ones who primarily isolated mesenchymal stem cells (MSCs) from bone marrow (BM).⁶ They were also isolated from other tissues such as adipose tissue,⁷ umbilical cord blood,⁸ endometrial tissue,⁹ and from dental pulp.¹⁰ MSCs have been used for regenerative purposes in patients.¹¹ Multi-lineage properties of MSCs into mesodermal and ectodermal lineages were previously demonstrated for osteoblasts,¹² neuronal-like cells,¹³ and heart muscles.¹⁴

Tissues such as bone marrow, adipose tissue, umbilical cord blood, placenta, dental pulp, and peripheral blood contain a pool of ASCs.^{15,16} Isolation and cultivation of these cells by standard protocols give the opportunity to use them in research and therapeutic application. The point is that the most isolation protocols are invasive and need to do surgical operation.^{17,18} Thus demands on finding an accessible source to harvest stem cells from an adult tissue through non-invasive methodology have increased.

The newly defined adult stem cells are menstrual blood-derived stem cells (MenSCs), giving rise to hopes in clinical application of these cells. They are mesenchymal-like stem cells that can be harvested from human menstrual blood shedding of endometrium monthly.^{19,20} MenSCs have a highly proliferation and differentiation capability under specific differentiation conditions.²¹ The easy and simple

way to get MenSCs without any invasive surgical intervention or hospitalization and absence of any ethical issues to isolate them are advantages of these MSCs.²²

Molecular profile assay shows that MenSCs express some pluripotency markers including Oct-4, SSEA-4, nanog, and c-kit and also some mesenchymal stem cells specific markers such as CD9, CD29, CD44.²³ So MenSCs are a good source of stem cells in research for differentiation into different cells and use in regenerative medicine. The differentiation of MenSCs into adipocytes, osteocytes, chondrocytes, hepatocytes, cardiomyocytes, and pancreatic cells has been previous demonstrated.²⁴

They can provide a new hope in regenerative medicine for their ability in differentiation into desired cells and tissues. Therefore, MenSCs would be a valuable choice in cell-based therapies and we can consider their potential in clinical trials especially in repair of dermatological lesions. Skin regeneration and repair has become the main goal of dermatological treatments including wrinkles, photoaging, cutaneous deep wounds, and burns as they are still major concerns in dermocosmetics.²⁵ So this study was conducted to isolate MenSCs and evaluate their potential in differentiation into epidermal lineage.

MATERIALS AND METHODS

The MenSCs were isolated from healthy fertile women aged between 22-30 years. All were volunteer donors giving a signed informed consent sheet according to ethical guideline of Avicenna Research Institute, Tehran, Iran. About 5-10 ml of menstrual blood (MB) was collected using sterile Diva cups inserted into vagina during menstruation. MB of Diva cups were then transferred into Falcon tube containing phosphate buffered saline (PBS) without Ca²⁺ or Mg²⁺ supplemented with 2.5 µg/ml fungizone, 100 µg/mL streptomycin, 100 U/mL penicillin and 0.5 mM EDTA.

Mononuclear cells were separated using Ficoll-Hypaque density gradient centrifugation and washed out in PBS. Then, the cell pellet in the tube was suspended in DMEM-F12 medium supplemented with 10% FBS and cultured in tissue culture plates. The cells were kept in 37°C incubator with 5% CO₂ and saturated humidity. After removal of non-adherent cells in second day of incubation, the culture of adherent cells

continued until 70% confluency.

To induce differentiation of isolated MenSCs into epidermal lineage, we co-cultured the isolated cells with keratinocytes derived from the foreskin of healthy newborn male aged 2-10 months who was a candidate for circumcision. We isolated the foreskin keratinocytes as described before.²⁶

RESULTS

The isolated MenSCs were adherent to the culture plates after 24 hours and easily explanted and highly proliferated. By reaching the 75% confluency, the cells exhibited spindle-shaped morphology like fibroblasts (Figure 1).

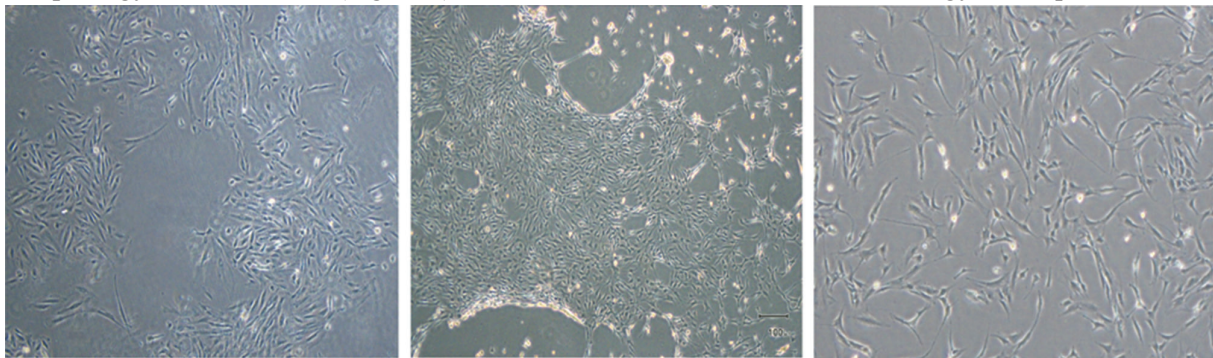


Fig. 1: Cultured menstrual blood-derived stem cells. The adherent cells showed spindle-shaped like morphology.

Flow cytometric analysis of cultured MenSCs revealed the expression of mesenchymal markers such as CD10, CD29, CD73, and CD105 (Figure 2). Further analysis showed the lack of hematopoietic stem cells markers (Figure 3). Under differentiation culture system, the spindle-shaped morphology of cultured MenSCs was changed into nearly irregular round to polygonal shape. At the end of the 2nd week, immunostaining assay of induced cells using indirect co-culture system revealed the expression of K14 and involucrin (Figure 4).

DISCUSSION

The stem cells technology has opened a new

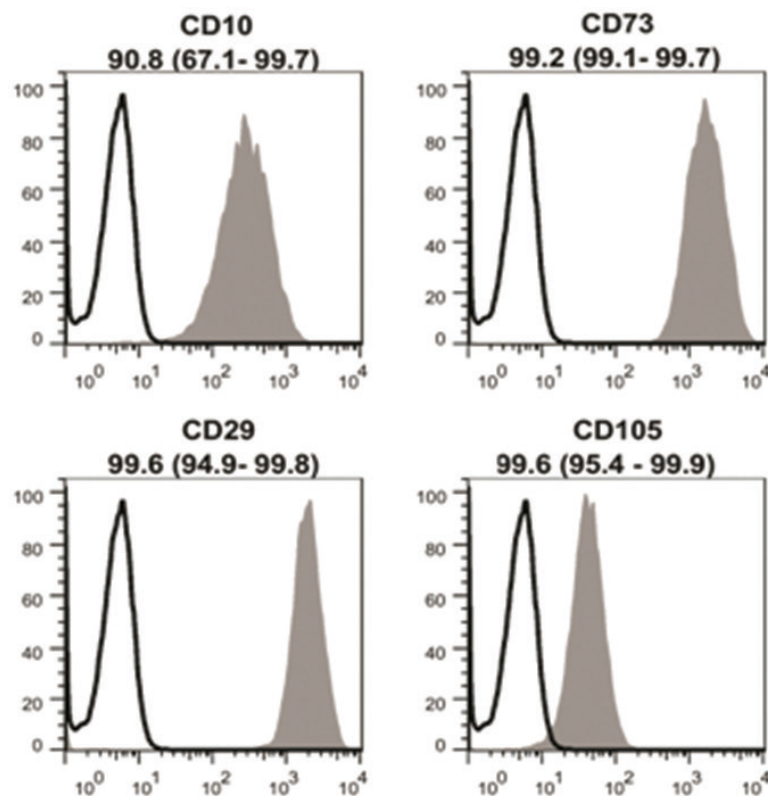


Fig. 2: Flowcytometric analysis of isolated menstrual blood-derived stem cells. The isolated cells expressed specific markers of mesenchymal stem cells.

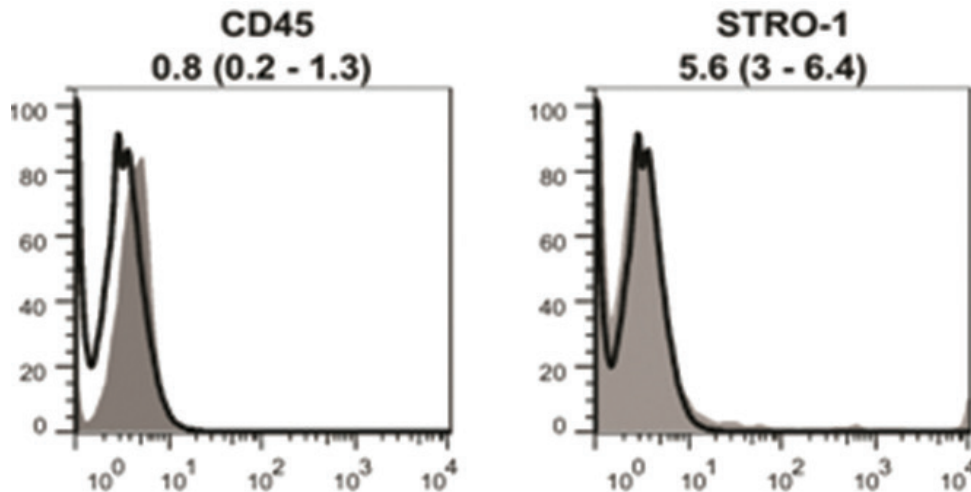


Fig. 3: Flowcytometric analysis of isolated menstrual blood-derived stem cells. The isolated cells did not express specific markers for hematopoietic stem cells.

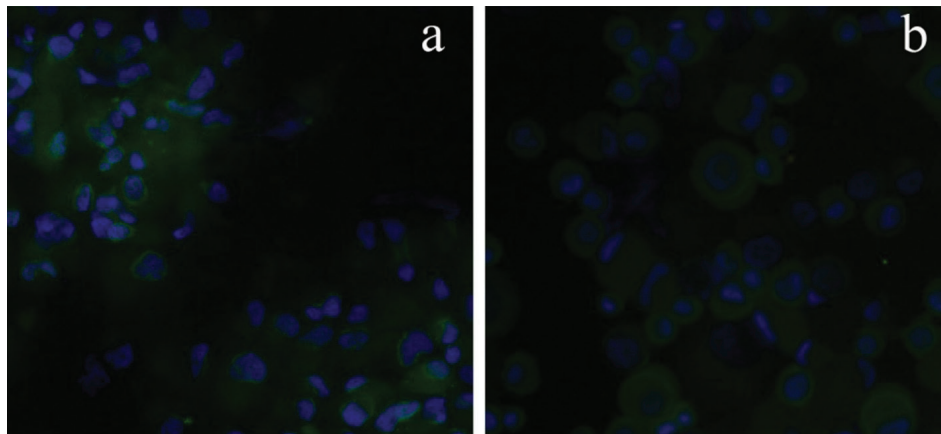


Fig. 4: Immunostaining of epidermal markers of involucrin (a) and K14 (b).

window to regenerative medicine.²⁷ In recent years many researchers attempted to find a safe, appropriate, and applicable way for regenerative purposes using stem cells²⁷ with specific characteristics of self-renewing and differentiating ability to repair damaged tissues.²⁸

Here, we designed a novel study to assess the differentiation potential of MenSCs into epidermal lineage for future repair of skin and dermatological lesions caused by ultraviolet rays, burn and chemicals damaging the integrity of skin tissue.²⁹ MSCs were shown to be an effective and attractive cell population in cell therapy to induce dermal repair and regeneration following acquired lesions and wounds.³⁰ They can provide essential trophic support to regenerate the injured tissue.

Kim et al. used adipose tissue derived mesenchymal stem cells (ADSCs) to eliminate UVB-induced wrinkles. This anti-wrinkled effect was mediated by decreasing UVB-

inducing apoptosis and stimulating collagen synthesis of dermal fibroblasts.³¹ Chen et al. reported the effect of cytokines and growth factors secreted by MSCs in regeneration of damaged skin tissue following full-thickness excisional wounds.³²

The newly defined mesenchymal-like stem cells from MB called MenSCs are a new source of stem cells³³ with good proliferation rate and capability in differentiation into various cell types similar to many other kinds of adult stem cells.²² Kazemnejad et al. investigated hepatic differentiation capacity of MenSCs compared to mesenchymal stem cells derived from bone marrow.²⁴ The derivation of adipogenic lineage, glial cells, and cardiogenic lineage were also demonstrated in other studies.³⁴⁻³⁶

In addition, these easily accessible adult stem cells have the capacity to trans-differentiate into neuronal cells, pancreatic cells, and osteocytes.³³ These investigations suggest this new source as

a safe alternative to other adult stem cells for cell therapies in different diseases. So we concluded the MenSCs could provide a suitable cell sources in repair and regenerate of skin diseases and naturally photoaging of skin. We are on the line to develop and suggest the standard inducing media and protocols to derive epidermal lineage from MenSCs. By achieving this goal, novel cell-based therapies will be proposed to use in many experimental and clinical studies.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- Shirazi R, Zarnani AH, Soleimani M, Abdolvahabi MA, Nayernia K, Ragerdi Kashani I. BMP4 can generate primordial germ cells from bone-marrow-derived pluripotent stem cells. *Cell Biol Int* 2012;**36**:1185-93.
- Alvarez CV, Garcia-Lavandeira M, Garcia-Rendueles ME, Diaz-Rodriguez E, Garcia-Rendueles AR, Perez-Romero S, Vila TV, Rodrigues JS, Lear PV, Bravo SB. Defining stem cell types: understanding the therapeutic potential of ESCs, ASCs, and iPS cells. *J Mol Endocrinol* 2012;**49**:R89-111.
- Gimond C, Marchetti S, Pagés G. Differentiation of Mouse Embryonic Stem Cells Into Endothelial Cells. In: *Embryonic Stem Cell Protocols* 2006; Springer, pp. 303-29.
- Shaterzadeh Yazdi H, Mehrabani D, Khodakaram Tafti A, Dianatpour M, Zare S, Tamaddon A, Razeghian Jahromi I. Osteogenic potential of subcutaneous adipose-derived stem cells in a rabbit model. *Onl J Vet Res* 2015;**19**:436-45.
- Ulrich D, Muralitharan R, Gargett CE. Toward the use of endometrial and menstrual blood mesenchymal stem cells for cell-based therapies. *Expert Opin Biol Ther* 2013;**13**:1387-400.
- Friedenstein AJ, Petrakova KV, Kurolesova AI, Frolova GP. Heterotopic of bone marrow. Analysis of precursor cells for osteogenic and hematopoietic tissues. *Transplantation* 1968;**6**:230-47.
- Mehrabani D, Mehrabani G, Zare S, Manafi A. Adipose-derived stem cells (ADSC) and aesthetic surgery: a mini review. *World J Plast Surg* 2013;**2**:65-70.
- Razmkhah F, Soleimani M, Mehrabani D, Karimi MH, Kafi-Abad SA. Leukemia cell microvesicles promote survival in umbilical cord blood hematopoietic stem cells. *EXCLI J* 2015;**14**:423-9.
- Mehrabani D, Rahmanifar F, Mellinejad M, Tamadon A, Dianatpour M, Zare S, Razeghian Jahromi I, Ghobadi F. Isolation, culture, characterization, and adipogenic differentiation of heifer endometrial mesenchymal stem cells. *Comp Clin Pathol* 2014 [Published on line]
- Mahdiyar P, Zare S, Robati R, Dianatpour M, Torabi K, Tamadon AD, Razeghian Jahromi I, Tamadon A, Mehrabani D. Isolation, Culture, and Characterization of Human Dental Pulp Mesenchymal Stem Cells. *Int J Pediatr 2nd Mashad Annual Stem Cell Research and Application Congress*. 2014.
- Ai J, Ebrahimi S, Khoshzaban A, Jafarzadeh Kashi TS, Mehrabani D. Tissue engineering using human mineralized bone xenograft and bone marrow mesenchymal stem cells allograft in healing of tibial fracture of experimental rabbit model. *Iran Red Crescent Med J* 2012;**14**:96-103.
- Ai J, Mehrabani D. The potential of human endometrial stem cells for osteoblast differentiation. *Iran Red Crescent Med J* 2010;**12**:585-7.
- Ai J, Noroozi Javidan A, Mehrabani D. The possibility of differentiation of human endometrial stem cells into neural cells. *Iran Red Crescent Med J* 2010;**12**:328-31.
- Ai J, Mehrabani D. Are endometrial stem cells novel tools against ischemic heart failure in women? a hypothesis. *Iran Red Crescent Med J* 2010;**12**:73-5.
- Aliborzi G, Vahdati A, Hossini SE, Mehrabani D. Evaluation of bone marrow-derived mesenchymal stem cells from Guinea pigs. *OnL J Vet Res* 2015;**19**:450-9.
- Secunda R, Vennila R, Mohanashankar AM, Rajasundari M, Jeswanth S, Surendran R. Isolation, expansion and characterisation of mesenchymal stem cells from human bone marrow, adipose tissue, umbilical cord blood and matrix: a comparative study. *Cytotechnology* 2014 May 6. [Epub ahead of print]
- Mehrabani D, Hassanshahi MA, Tamadon A, Zare S, Keshavarz S, Rahmanifar F, Dianatpour M, Khodabandeh Z, Jahromi I,

- Tanideh N, Ramzi M, Aqababa Hr, Kuhi-Hoseinabadi O. Adipose tissue-derived mesenchymal stem cells repair germinal cells of seminiferous tubules of busulfan-induced azoospermic rats. *J Hum Reprod Sci* 2015;**8**:103-10.
- 18 Isakson M, de Blacam C, Whelan D, McArdle A, Clover AJ. Mesenchymal stem cells and cutaneous wound healing: current evidence and future potential. *Stem Cells Int* 2015;**2015**:831095.
 - 19 Ghobadi F, Mehrabani D, Mehrabani G. Regenerative potential of endometrial stem cells: a mini review. *World J Plast Surg* 2015;**4**:1-6.
 - 20 Patel AN, Park E, Kuzman M, Benetti F, Silva FJ, Allickson JG. Multipotent menstrual blood stromal stem cells: isolation, characterization, and differentiation. *Cell Transplant* 2008;**17**:303-11.
 - 21 Kazemnejad S, Akhondi MM, Soleimani M, Zarnani AH, Khanmohammadi M, Darzi S, Alimoghadam K. Characterization and chondrogenic differentiation of menstrual blood-derived stem cells on a nanofibrous scaffold. *Int J Artif Organs* 2012;**35**:55-66.
 - 22 Allickson JG1, Sanchez A, Yefimenko N, Borlongan CV, Sanberg PR. Recent studies assessing the proliferative capability of a novel adult stem cell identified in menstrual blood. *Open Stem Cell J* 2011;**3**:4-10.
 - 23 Mou XZ, Lin J, Chen JY, Li YF, Wu XX, Xiang BY, Li CY, Ma JM, Xiang C. Menstrual blood-derived mesenchymal stem cells differentiate into functional hepatocyte-like cells. *J Zhejiang Univ Sci B* 2013;**14**:961-72.
 - 24 Khanjani S, Khanmohammadi M, Zarnani AH, Akhondi MM, Ahani A, Ghaempanah Z, Naderi MM, Eghtesad S, Kazemnejad S. Comparative evaluation of differentiation potential of menstrual blood-versus bone marrow-derived stem cells into hepatocyte-like cells. *PLoS One* 2014;**9**:e86075.
 - 25 Yildirim L, Thanh NT, Seifalian AM. Skin regeneration scaffolds: a multimodal bottom-up approach. *Trends Biotechnol* 2012;**30**:638-48.
 - 26 Zare S, Zarei MA, Ghadimi T, Fathi F, Jalili A, Hakhamaneshi MS. Isolation, cultivation and transfection of human keratinocytes. *Cell Biol Int* 2014;**38**:444-51.
 - 27 Hosseinkhani M, Mehrabani D, Karimfar MH, Bakhtiyari S, Manafi A, Shirazi R. Tissue engineered scaffolds in regenerative medicine. *World J Plast Surg* 2014;**3**:3-7.
 - 28 Kashani IR, Golipoor Z, Akbari M, Mahmoudi R, Azari S, Shirazi R, Bayat M, Ghasemi S. Basic research Schwann-like cell differentiation from rat bone marrow stem cells. *Arch Med Sci* 2011;**7**:45-52.
 - 29 Kim AY, Lee EM, Lee EJ, Min CW, Kang KK, Park JK, Hong IH, Ishigami A, Tremblay JP, Jeong KS. The wound-healing and antioxidant effects of adipose-derived stem cells. *Cell Transplant* 2013;**22**:1845-58.
 - 30 Yolanda MM, Maria AV, Amaia FG, Marcos PB, Silvia PL, Dolores E, Jesús OH. Adult stem cell therapy in chronic wound healing. *J Stem Cell Res Ther* 2014;**4**:162.
 - 31 Kim WS, Park BS, Park SH, Kim HK, Sung JH. Antiwrinkle effect of adipose-derived stem cell: activation of dermal fibroblast by secretory factors. *J Dermatol Sci* 2009;**53**:96-102.
 - 32 Chen L, Tredget EE, Wu PY, Wu Y. Paracrine factors of mesenchymal stem cells recruit macrophages and endothelial lineage cells and enhance wound healing. *PLoS One* 2008;**3**:e1886.
 - 33 Meng X, Ichim TE, Zhong J, Rogers A, Yin Z, Jackson J, Wang H, Ge W, Bogin V, Chan KW, Thébaud B, Riordan NH. Endometrial regenerative cells: a novel stem cell population. *J Transl Med* 2007;**5**:57.
 - 34 Khanmohammadi M, Khanjani S, Edalatkhah H, Zarnani AH, Heidari-Vala H, Soleimani M, Alimoghaddam K, Kazemnejad S. Modified protocol for improvement of differentiation potential of menstrual blood-derived stem cells into adipogenic lineage. *Cell Prolif* 2014;**47**:615-23.
 - 35 Rahimi M, Zarnani AH, Mohseni-Kouchesfehani H, Soltanghoraei H, Akhondi MM, Kazemnejad S. Comparative Evaluation of Cardiac Markers in Differentiated Cells from Menstrual Blood and Bone Marrow-Derived Stem Cells In Vitro. *Mol Biotechnol* 2014;**56**:1151-62.
 - 36 Azedi F, Kazemnejad S, Zarnani AH, Behzadi G, Vasei M, Khanmohammadi M, Khanjani S, Edalatkhah H, Lakpour N. Differentiation potential of menstrual blood-versus bone marrow-stem cells into glial-like cells. *Cell Biol Int* 2014;**38**:615-24.